

## In this guide:

- Recommended settings for the BioTek Synergy<sup>™</sup> 2 and 4 Multi-Mode Microplate Readers using the Transcreener ADP<sup>2</sup> Assay in 96-Well format
- Optimizing sensitivity setting and flash number improves data
- Synergy<sup>™</sup> 2 and 4 meet requirements for BellBrook Labs' Instrument Validation Program when used with the Standard PMT detector in 96-Well format



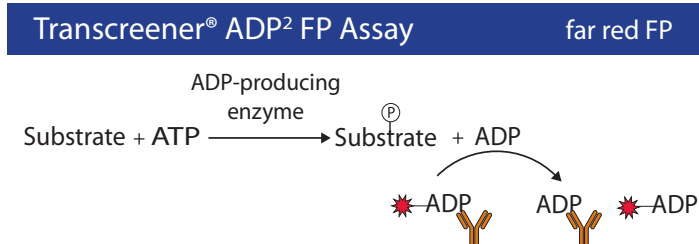
## Introduction

Transcreener is a universal, high throughput biochemical assay based on detection of nucleotides, which are formed by thousands of cellular enzymes - many of which catalyze the covalent regulatory reactions that are central to cell signaling and are high value targets in drug discovery. The advantages of the Transcreener<sup>®</sup> HTS Assay Platform over existing assay methods include the following. **Universality**-The detection of invariant nucleotide reaction product means that a single set of detection reagents can be used for all of the enzymes in a family and all acceptor substrates. **Far Red Fluorescence Polarization Detection**-Use of far red shifted dyes with a ratiometric output greatly reduces interference and particulate-based light scattering from fluorescent compounds. **Sensitivity**-High affinity antibodies enable robust detection of low levels of substrate conversion (<10%) with less enzyme than other methods.

A critical factor in realizing the numerous advantages of the Transcreener HTS assays is the correct setup of the microplate reader. Proper selection of filters, dichroics, monochromator settings, and read times impact an instrument's sensitivity with any given assay. In response to this fact, BellBrook Labs has developed an Instrument Validation Program to test and optimize plate readers. This will ensure that researchers are aware of the readers meeting minimal performance requirements, as well as knowing the most optimal settings for each detection system.

## Assay Principle

BellBrook Labs' Instrument Validation Program employs the Transcreener<sup>®</sup> ADP<sup>2</sup> Assay for all testing. The excitation peak is centered at 633nm, while the emission peak centers at 650nm. The Transcreener ADP<sup>2</sup> assay is a simple one-step homogenous detection assay. Transcreener ADP Detection Mixture, comprised of an ADP Alexa633 Tracer bound to an ADP<sup>2</sup> Antibody, is added to an equal volume of enzyme reaction mix. Enzymatically generated ADP displaces the tracer resulting in a decrease in fluorescence polarization. Standard curves are generated using varying concentrations of ATP and ADP to mimic the conversion of ATP to ADP during the course of an enzyme reaction.



## Validation Criteria

- 96-Well Format
- Z'-Factor  $\geq 0.7$  at 10% conversion of 10  $\mu\text{M}$  ATP
- $\Delta$  mP  $\geq 95$  mP at 10% conversion of 10  $\mu\text{M}$  ATP
- Z' and  $\Delta$  mP specifications to be met using Transcreener<sup>®</sup> ADP<sup>2</sup> Assay reagents
- Read time to achieve Z' and  $\Delta$  mP specifications  $\leq 5$  minutes

The results from the validation using the Transcreener ADP<sup>2</sup> Assay are representative of the performance with other Transcreener Far Red FP Assays with this instrument in 96-well format. Because individual results may vary, optimal setting recommendations should be used as a starting point. Slight modifications may be necessary to optimize results for your particular assay on your instrument.

## Synergy<sup>™</sup> 2 and Synergy<sup>™</sup> 4 Information

### Synergy<sup>™</sup> 2



- Detection modes available as individual modules
- Plate formats: 6- to 1536-well plates
- Unique combination of monochromator, filters and dichroic mirrors for best performance in all modes
- 3 broad spectrum light sources for optimal sample illumination
- Powered by Gen5 Data Analysis Software

### Synergy<sup>™</sup> 4



- Hybrid Technology combines high-performance filter system with flexible monochromator system
- Detection modes available as individual modules
- Plate formats: 6- to 1536-well plates
- 3 broad spectrum light sources for optimal sample illumination
- Powered by Gen5 Data Analysis Software

## Materials and Methods

**Instrument:** Synergy™ 2 Multi-Mode Microplate Reader

**Microplates:** Corning® 96 Well Half Area Black Flat Bottom PS NBS™ Microplate (Product #3686)

### Reagents

Reagent	Kit/Component Catalog #
Transcreener® ADP <sup>2</sup> FP Assay	3010-1K
ADP Alexa633 Tracer, 400 nM	2009
Stop & Detect Buffer B, 10X	2027
ADP <sup>2</sup> Antibody	2051
5 mM ADP	2052
5 mM ATP	2053
Buffer Components	
500 mM EGTA	Not Provided
1000 mM HEPES	
500 mM MgCl <sub>2</sub>	
1% Brij-35	
100% DMSO	

**Table 1. Experimental Reagents**

### Protocol

Standard protocol consists of adding 30 µL of ADP Detection Mixture to 30 µL of the ATP/ADP Standard Mixture in a 96 well plate. The plate was then covered, shaken to mix the reagents, and incubated at room temperature for 60 minutes.

### ATP/ADP Mixture

The ATP/ADP mixture consists of 4 mM MgCl<sub>2</sub>, 2 mM EGTA, 50 mM HEPES, pH 7.5, 1% DMSO, 0.01% Brij-35, and ATP/ADP combined to a constant adenine concentration of 10 µM.

### ADP Detection Mixture

The ADP Detection Mixture consists of 1X Stop & Detect Buffer B, 4 nM ADP Alexa633 Tracer, and 14.8 µg/mL ADP<sup>2</sup> Antibody.

### Free Tracer

The Free Tracer consists of 1X Stop & Detect Buffer B, and 4 nM ADP Alexa633 Tracer.

### Buffer Blank

The Buffer Blank consists of 1X Stop & Detect Buffer B, and 14.8 µg/mL ADP<sup>2</sup> Antibody.

### Final Concentrations in 60 µL Reaction Volume

2 mM MgCl<sub>2</sub>, 1 mM EGTA, 25 mM HEPES (pH 7.5), 0.5% DMSO, 0.005% Brij-35, ATP/ADP combined to a constant adenine concentration of 5 µM, 0.5X Stop & Detect Buffer B (25 mM HEPES, pH 7.5, 200 mM NaCl, 10 mM EDTA, and 0.01% Brij-35), 2 nM ADP Alexa633 Tracer, and 7.4 µg/mL ADP<sup>2</sup> Antibody.

### Standard Curve Preparation

8-point ATP/ADP standard curves were generated to test the Synergy™ 2 Microplate Reader. ATP/ADP mixtures were created at the various concentrations of ATP and ADP listed in Table 2. Final concentration of the buffer components are listed above. Each point on the curve mimics a different substrate conversion level in an enzyme reaction (n=11). 30 µL of each ATP/ADP combination was dispensed to 11 wells of an individual row of a 96-well plate. 30 µL of the 10 µM ATP/0 µM ADP combination was also dispensed to wells A12 and B12 of the plate.

30 µL of the prepared ADP Detection Mixture was then dispensed to wells A1-H11 of the assay plate. Finally, in place of the ADP Detection Mixture, 30 µL of free tracer was dispensed to well A12, and 30 µL of buffer blank was dispensed to well B12.

Substrate Conversion Levels (%)	ATP, µM	ADP, µM
0	10	0
4	9.6	0.4
8	9.2	0.8
10	9	1
15	8.5	1.5
30	7	3
60	4	6
100	0	10

**Table 2. Standard Curve ATP/ADP Concentrations**

### Instrument Set-up and Filter Information

Instrument Detection Components		BioTek Catalog #
Excitation Filter	620/40 nm	7082213
Emission Filter	680/30 nm	7082229
Dichroic	660 nm cutoff	7137660
PMT	Standard PMT detector	49984

**Table 3. Instrument Optics**

The excitation and emission filters, dichroic mirror, and Standard PMT were installed prior to instrument evaluation. Following the installation process, and prior to the first read of a plate, the following steps were taken to optimize the detector gain with the use of the Sensitivity setting, which prepared the Synergy™ 2 Microplate Reader to correctly read the Transcreener® ADP<sup>2</sup> FP Assay:

1. The Read Step was opened within the Procedure to be used to read the plate.
2. The Sensitivity “Options” was chosen, and “Automatic Sensitivity Adjustment” and “Scale to High Wells” were selected.
3. A well containing free tracer was chosen, and the “Scale Value” was set to 50,000.

This function allows the user to select a well or a group of wells on the plate, and have the reader scale the raw measurement to these wells. The recommended “Scale Value” is 50,000 RFU. This value allows for the use of the full measurement range of the system, while leaving some room for samples that might be a little brighter than expected. Typically, the wells used for the scaling are positive controls, the highest standards of the standard curve, or any other well expected to provide a strong signal. Subsequent reads using the same plate used the predetermined sensitivity settings. To ascertain this value the following steps were completed:

1. The Procedure options were opened by right clicking on “Procedure”.
2. “AutoSensitivity Results” was chosen. The sensitivity setting was recorded from the “AutoSensitivity Results” screen.
3. The Read Step was once again opened, and the Sensitivity Options was chosen.
4. The “Automatic Sensitivity Adjustment” option was deselected. This allowed the value used for the initial read to be entered into the “Sensitivity” box.

Optimized Measurement Settings	
Sensitivity Setting	160
Top Probe Vertical Offset	5.00
Flash Number	variable

**Table 4. Instrument Settings**

Fluorescence polarization measurements were performed using the settings listed in Table 4. The sensitivity setting was optimized prior to reading the plate, using the procedure previously described. The number of flashes per well and top probe vertical offset were manually adjusted in the Read Step of the Protocol. Flash number was varied to determine the range of read times that would meet the criteria of the instrument validation program.

### Calculations

#### Δ mP Calculation

mP values for each substrate percent conversion level were subtracted from the mP value at 0% ATP conversion.

$$mP = mP_{\text{initial [ATP]}} - mP_{\text{sample}}$$

The change in mP values (Δ mP) is indicative of the amount of ATP that is converted to ADP in an enzyme reaction. A Δ mP of approximately 100 mP units is ideal in a compound screening situation.

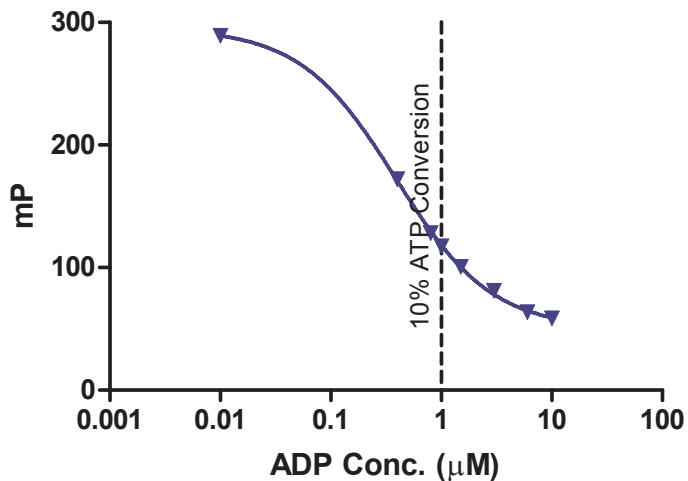
#### Z'-Factor Calculation

$$Z' = 1 - [(3 * SD_{\text{initial [ATP]}} + 3 * SD_{\text{sample}}) / (mP_{\text{initial [ATP]}} - mP_{\text{sample}})]$$

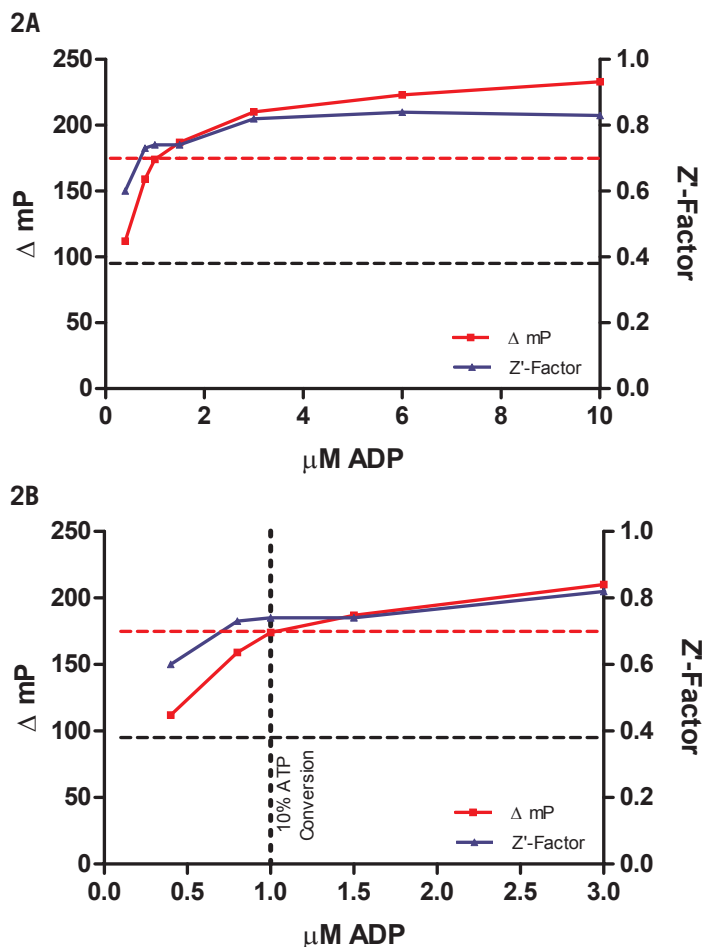
While an assay yielding a  $Z' \geq 0.5$  is considered a high quality assay, those producing  $Z'$  values  $\geq 0.7$  give the user a greater confidence level.

## Results

Assay plates containing the 8-point standard curve were read on the Synergy™ 2 Microplate Reader (Figure 1). As the ratio of ADP:ATP increases the proportion of bound tracer vs. free tracer decreases, resulting in an overall decrease in mP values.



**Figure 1. 10 μM ATP/ADP Standard Curve. 10% ATP Conversion represents 9 μM ATP/1 μM ADP concentration level.**



**Figure 2. A)  $Z'$  and  $\Delta$  mP values observed in a standard curve mimicking conversion of 10 μM ATP to ADP. B) A zoom in of the 1-3 μM ADP section of the standard curve.  $Z'$  validation minimal qualification shown by red dashed line.  $\Delta$  mP validation minimal qualification shown by black dashed line. 10% ATP conversion validation point shown by black dotted line. Reader set at 15 flashes.**

A  $Z' > 0.7$  and an mP shift of 174 units is achieved at 1.0 μM ADP (10% ATP conversion) in a read time of 1:39 (Figure 2A). The  $Z'$  value falls below 0.7 at 4% substrate conversion, which coincides with a Δ mP value of 112. Figure 2B highlights data that is generated in the initial velocity range of the reaction.

Assay Performance at 10% Conversion of 10 μM ATP			
Flashes	15	20	25
Read Time (Minutes)	1:39	1:59	2:19
10% ATP Conversion ΔmP	174	174	172
10% ATP Conversion Std. Dev.	10	9	6
10% ATP Conversion $Z'$ -Factor	0.74	0.71	0.80

**Table 5. Assay performance with various instrument settings.**

Variable flash numbers were evaluated to determine the optimal read time generating the highest quality data. As flash number increases, the standard deviation of the mean FP values decreases, resulting in improved  $Z'$  values (Table 5). The shortest plate read that results in data meeting validation criteria occurs with 15 flashes.

## Discussion

The data shows that the BioTek Synergy™ 2 and 4 are compatible with the Transcreener Far Red FP Detection Module in 96-well format. Following the Transcreener protocol (using 30 µL volumes instead of 10 µL), and setting the instrument to 15 flash reads or above yielded Z' values  $\geq 0.7$ . The data also shows that the Transcreener ADP<sup>2</sup> FP Assay yields excellent data quality at low % ATP conversion levels, thereby making it possible to use smaller amounts of enzyme and substrate in an enzyme reaction. It is important to use the instrument setup described in the materials and methods. A change in filters or dichroics may have adverse effects on instrument performance resulting in an increased standard error in reads.

## Conclusions

- BioTek Synergy™ 2 and 4 Microplate Readers passed the 96-well validation criteria under the following conditions: Using a 620/40 nm excitation filter, a 680/30 nm emission filter, a 660 nm cutoff dichroic, and the Standard PMT detector; instrument was set to 15 flashes and yielded a Z' > 0.7 in 1:39 minutes
- Using optimized instrument settings and the filters and dichroic recommended by BioTek reduces standard error in mP reads
- Transcreener® ADP<sup>2</sup> FP Assay improves data quality using initial rate enzyme reaction kinetics

## Additional Information

### Related Products

Transcreener® ADP <sup>2</sup> FP Assay.....	3010-1K
Transcreener® ADP <sup>2</sup> TR-FRET Red Assay.....	3011-1K
Transcreener® ADP <sup>2</sup> FI Assay.....	3013-1K
Transcreener® AMP/GMP Assay.....	3006-1K
Transcreener® UDP Assay.....	3007-1K
Transcreener® GDP FP Assay.....	3009-1K

### Ordering Information

Please contact BellBrook Labs for product pricing. Custom quotes are available for orders of 10,000 wells or more.

Phone orders:  
608.443.2400  
866.313.7881

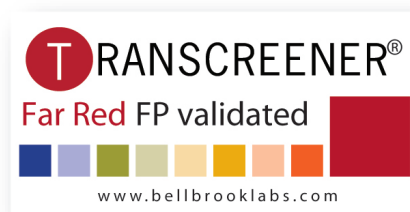
Fax orders:  
608.441.2967

Email orders:  
info@bellbrooklabs.com

### Technical Information

For technical information, please contact Meera Kumar, Applications Scientist  
608.443.2400  
866.313.7881  
meera.kumar@bellbrooklabs.com

### Transcreener Instrument Validation Stickers



Look for the Transcreener Far Red FP-validated sticker on instruments that have successfully met our validation criteria.

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